VAHTS RNA Clean Beads

N412

Version 22.1

Product Description

Vazyme VAHTS RNA Clean Beads is based on SPRI (Solid Phase Reverse Immobilization) and is applicable for RNA purification by efficiently removing all proteins, salt ions, and other impurities. The usage of VAHTS RNA Clean Beads is the same as the same products of other suppliers.

Components

Components	N412-01	N412-02	N412-03
VAHTS RNA Clean Beads	5 ml	40 ml	450 ml

Storage

Store at 2 ~ 8℃. Adjust the shipping method according to the destination.

Applications

It is applicable for RNA purification from in vitro reaction mixtures, i.e. RNA library preparation, and not applicable for direct RNA extraction from cells or tissues.

Self-prepared Materials

Ethanol (100%)
Nuclease-free ddH_2O and tubes
Magnetic stand

Notes

For research use only. Not for use in diagnostic procedures.

1. Keep the VAHTS RNA Clean Beads at room temperature at least 30 min and shake the reagent well before use, otherwise the recovery efficiency of the sample should be affected.
2. RNase and nucleic acid contamination must be avoid during the experiment.
3. The 80% ethanol should be prepared with Nuclease-free ddH_2O to avoid RNA degradation.
4. The drying time should be controlled to ensure there is no residual ethanol and avoid excessive drying, which may cause cracking on the surface of beads and thereby reduce the recovery of RNA.
5. In the protocol Step 10, avoid pipetting beads when transferring the supernatant, i.e. leave behind 2 - 3 μl of supernatant.
6. The VAHTS RNA Clean Beads is compatible for various library prep kits, such as VAHTS Universal V8 RNA-seq Library Prep Kit for Illumina (Vazyme #NR605). Please refer to the protocols in these kits when using the VAHTS RNA Clean Beads for library preparation.

Experiment Process

1. Take out the VAHTS RNA Clean Beads from 2 ~ 8℃ and keep the reagent at room temperature at least 30 min before use.
2. Mix the VAHTS RNA Clean Beads thoroughly by vortex or turning upside down. Add VAHTS RNA Clean Beads into the RNA solution with volume = 1.8 × volume of RNA solution.
3. Mix the VAHTS RNA Clean Beads and sample thoroughly by pipette mixing 10 times.
4. Incubate the tube at room temperature for 5 min for the binding of RNA to beads.
5. Place the tube onto the magnetic stand for 5 min. Until the solution becomes clear, carefully pipette the supernatant from the tube and discard.
6. Keep the tube on magnetic stand. Dispense 200 μl of freshly prepared 80% ethanol into the tube and incubate for 30 sec at room temperature. DO NOT re-suspend the beads! Pipette out the ethanol and discard.
7. Repeat the step 6.
8. Keep the tube on the magnetic stand and uncap the tube to air-dry the beads for 5 - 10 min.
9. Take out the tube from magnetic stand. Add an appropriate amount of Nuclease-free ddH_2O to the tube, manually resuspend the beads by pipetting up and down 10 times, and then incubated at room temperature for 5 min.
10. Place the tube to magnetic stand for 5 min. After the solution to be clarified, pipette out the supernatant to a new Nuclease-free centrifuge tube without disturbing the beads.
11. Store the supernatant at -30 ~ -15℃ or proceed to the next step immediately.